

High-throughput Single-cell Targeted DNA Sequencing Using an Updated Tapestri™ Platform Reveals Rare Clones and Clonal Evolution for Multiple Blood Cancers

AACR-4696

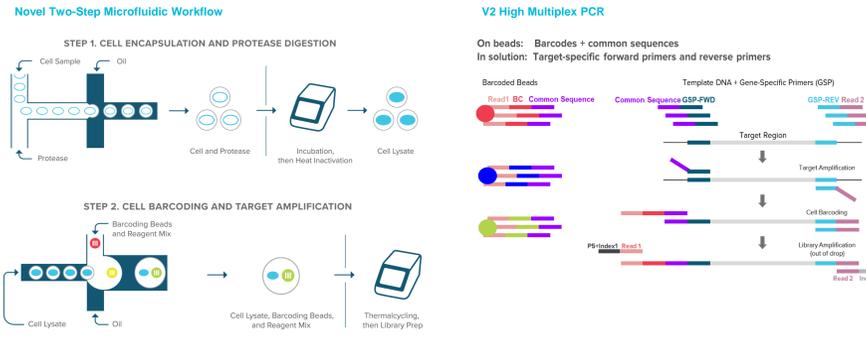
Nianzhen Li, Daniel Mendoza, Adam Sciambi, Mani Manivannan, Jacob Ho, Kaustubh Gokhale, Benchun Liu, Jacqueline Marin, Kathryn Thompson, Jamie Yates, Vasu Sharma, Steven Chow, Sombeet Sahu, Shu Wang, Dennis Eastburn, Keith Jones, Nigel Beard
Mission Bio, Inc., South San Francisco, CA, USA



Abstract

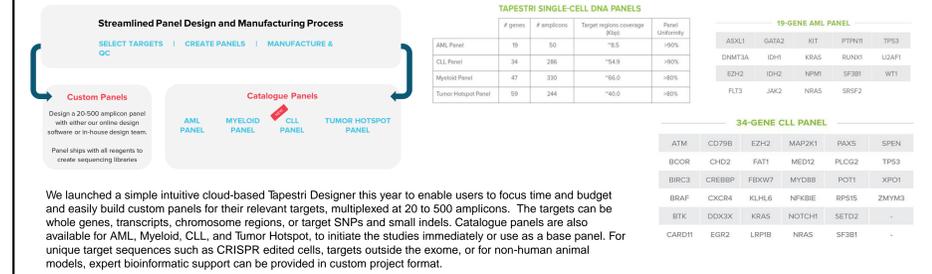
The challenge in precision medicine has been improving the understanding of cancer heterogeneity and clonal evolution, which has major implications in targeted therapy selection and disease monitoring. However, current bulk sequencing methods are unable to unambiguously identify rare pathogenic or drug-resistant cell populations and determine whether mutations co-occur within the same cell. Single-cell sequencing has the potential to provide unique insights on the cellular and genetic composition, drivers, and signatures of cancer at unparalleled sensitivity. **Methods:** Previously we have developed a high-throughput single-cell DNA analysis platform (Tapestri™) that leverages droplet microfluidics and a multiplex-PCR based targeted DNA sequencing approach, and demonstrated the generation of high-resolution maps of clonal architecture from acute myeloid leukemia (AML) tumors. Here we present an update to the Tapestri Platform which employs new biochemistry and features improved firmware, software, workflow, and data analysis solution resulting in higher throughput, better sensitivity, specificity and unprecedented flexibility. **Results:** From cell prep to sequencing-ready libraries, the workflow can be completed within 2 days. We have validated the performance of an AML (19 genes, 50 amplicons) and a CLL (chronic lymphocytic leukemia) (34 genes, 286 amplicons) panel. We also developed a robust web-based design portal for custom targets. The updated biochemistry enables easy addition of new gene and loci targets into existing panels for improved coverage and updated studies. Using longitudinal AML and CLL samples, we were able to detect rare subclones of <0.1% prevalence, identify mutation co-occurrence, and characterize clonal evolution due to disease progression and drug treatment. **Conclusion:** We demonstrate that single-cell DNA sequencing can reveal the heterogeneity of blood cancers and map the clonal architecture and clonal evolution with higher sensitivity than bulk NGS methods. This is critical in patient stratification and drug selection over the entire course of treatment. Besides the catalog AML and CLL panels, the flexibility of system allows for analyzing SNV and indel mutations of any custom cancer DNA targets. Additionally, the system provides capabilities for quality control of gene edited cells, further advancing research into cancer therapies.

The Foundations of Tapestri Technology

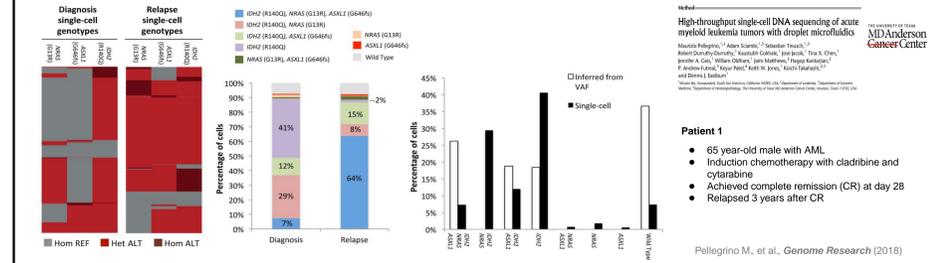


The Tapestri precision genomic platform is enabled by a novel two-step microfluidic workflow and a high multiplex PCR biochemistry scheme. The two-step microfluidics allows for efficient access to DNA for downstream genomic reactions and provides flexibility to adapt for additional applications and multi-omics. The multiplex PCR chemistry is developed and co-optimized with an AI-powered panel design pipeline and enables direct and efficient amplification of targeted genomic regions within barcoded individual cells. Taken together the platform produces high genomic coverage, low allele dropout rate, highly uniform amplification in thousands of cells from single run, is compatible with diverse and difficult samples, and is easily deployable for custom content.

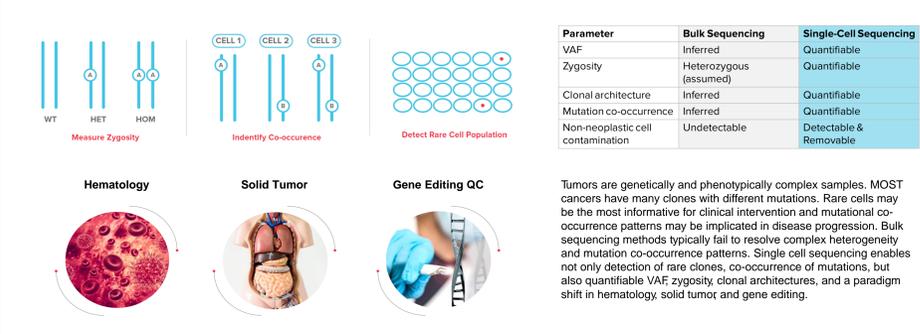
Build Desired Targeted Panels



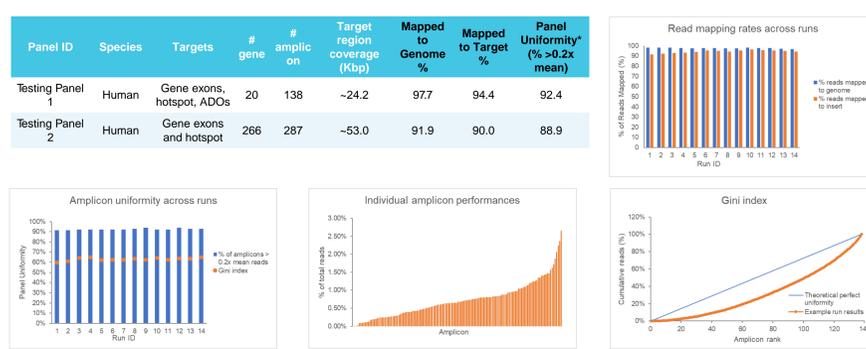
Clonal Evolution in Longitudinal AML Samples



The Power of Single Cell DNA Analysis

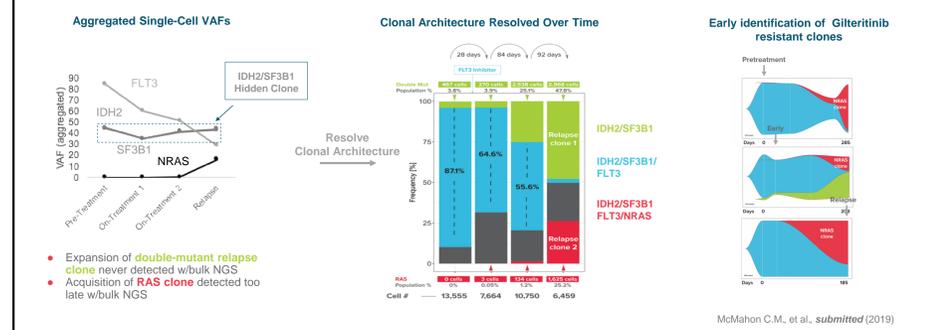


Performances of Testing Panels

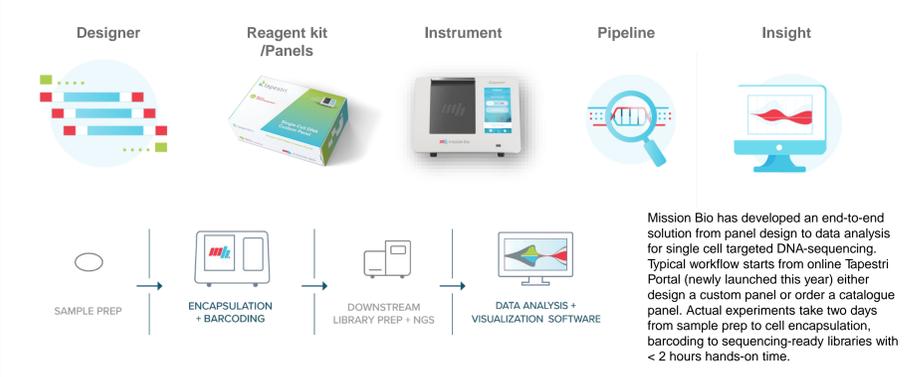


To test the performances of the updated chemistry and designer pipeline, we designed three panels with 31, 138, and 287 (small, medium, large) amplicons respectively. The primer pools were pooled in equal volumes without any optimization, in order to simulate a standard custom panel at first pass (Mission Bio can also provide white-glove services to rescue specific designs and optimize individual amplicon performances after wet QC). Multiple runs were conducted for each panel with different cell types. An updated Tapestri analysis pipeline was used in analyzing the FASTQ data, collapsing cell barcodes, finding cells and calling genotypes. Typical cell outputs range from 5000 to 7,000 cells in these runs (30-40x sequencing depth), with <8% cell mixing (50/50 cell mix experiments) and <10% allele dropout rate. The run to run consistency data were shown above from multiple runs of Testing Panel 1. Bottom graphs show the panel uniformity for the runs and how the panel uniformity metrics were calculated and visualized.

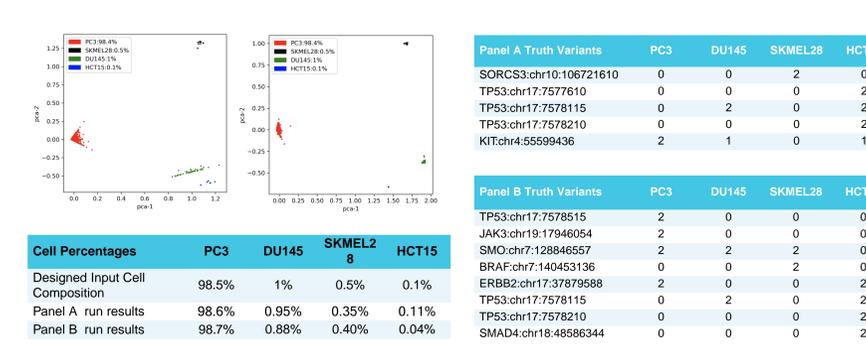
Tapestri Reveals Therapy Resistant Clones



Tapestri Precision Genomics Platform Overview

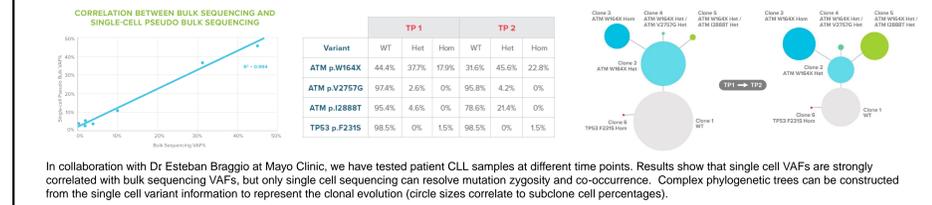


High Sensitivity Detection of Rare Cell Populations



To test the sensitivity and specificity of the system, we mixed four types of cells at different concentrations (98.4%, 1%, 0.5%, and 0.1%), and run the mixed cell population on Tapestri with two targeted panels (Panel A-50 amplicons and Panel B-244 amplicons). The truth variant data from the four cell types were generated from bulk sequencing of pure cell lines, then used in Tapestri data analysis to determine cell types for all single cells from the Tapestri results. The resulted percentages matched closely to the desired cell input composition. Clones of 0.1% cells can be detected using both panels.

Single-Cell DNA Sequencing Resolves the Genetic Complexity Underlying CLL Progression



Conclusions

The Tapestri Precision Genomics Platform utilizes novel droplet microfluidics to access the genomic DNA. Together with updated chemistry and designer it generates high coverage uniformity and low allele dropout for thousands of single cells, and enables detection of rare subclones down to 0.1%. Researchers now have a highly sensitive, targeted, and customizable solution for investigating genomic variation and clonal evolution in complex biological samples, as demonstrated in multiple AML and CLL longitudinal studies.

References:
Pellegrino M, et al., *Genome Research* (2018)
McMahon C.M., et al., *submitted* (2019)

Single Cell DNA-Seq Data Analysis and Visualization

