## Abstract #1663

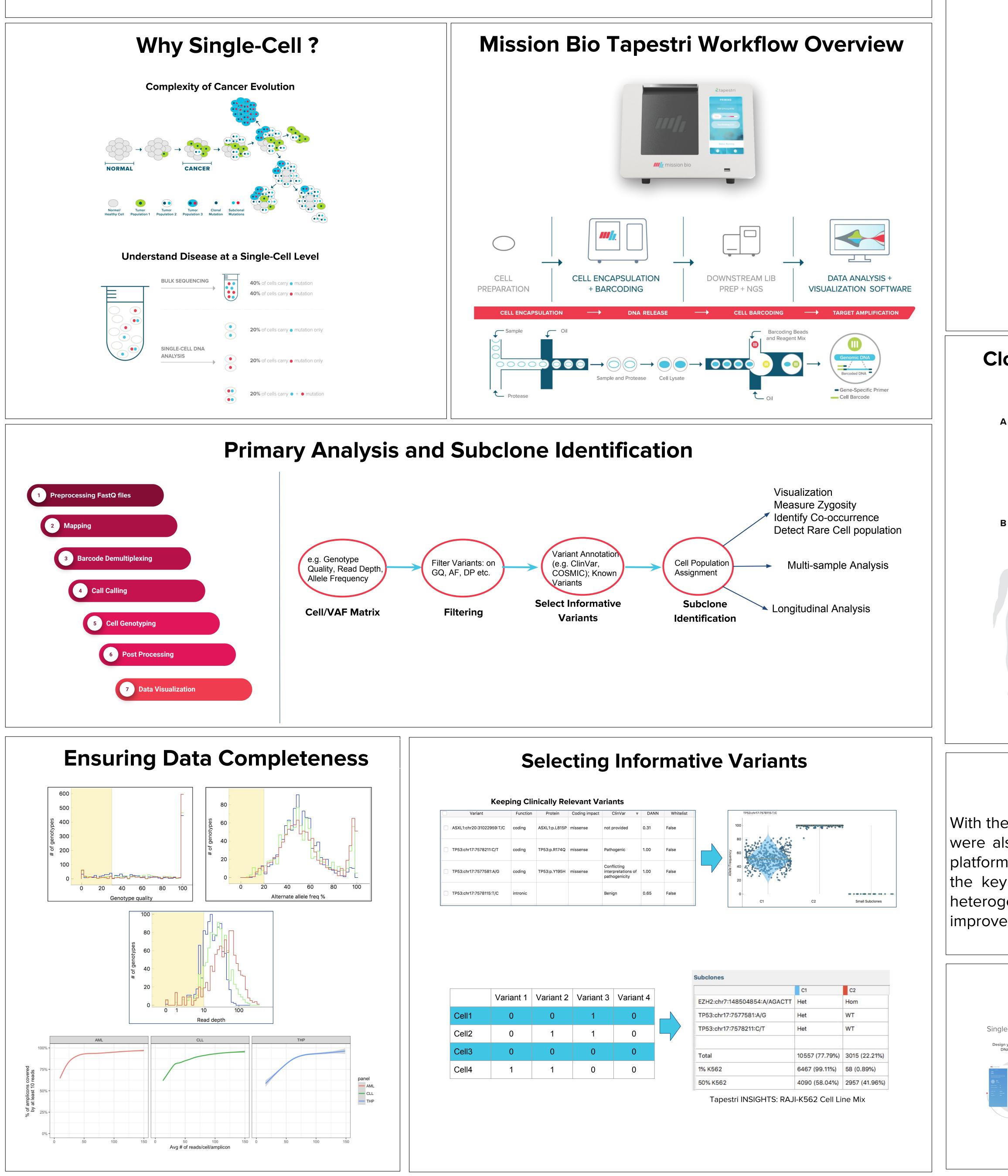
# **Analytical Methods to Identify Tumor Heterogeneity and Rare** Subclones in Single Cell DNA Sequencing Data from Targeted Panels Manimozhi Manivannan<sup>1</sup>, Sombeet Sahu<sup>1</sup>, Shu Wang<sup>1</sup>, Dong Kim<sup>1</sup>, Niranjan Vissa<sup>1</sup>, Kaustubh Gokhale<sup>1</sup>, Adam Sciambi<sup>1</sup>, Nianzhen Li<sup>1</sup>, Robert Durruthy-Durruthy<sup>1</sup>, Anup Parikh<sup>1</sup>, Hannah Viernes<sup>1</sup>, Keith Jones<sup>1</sup>

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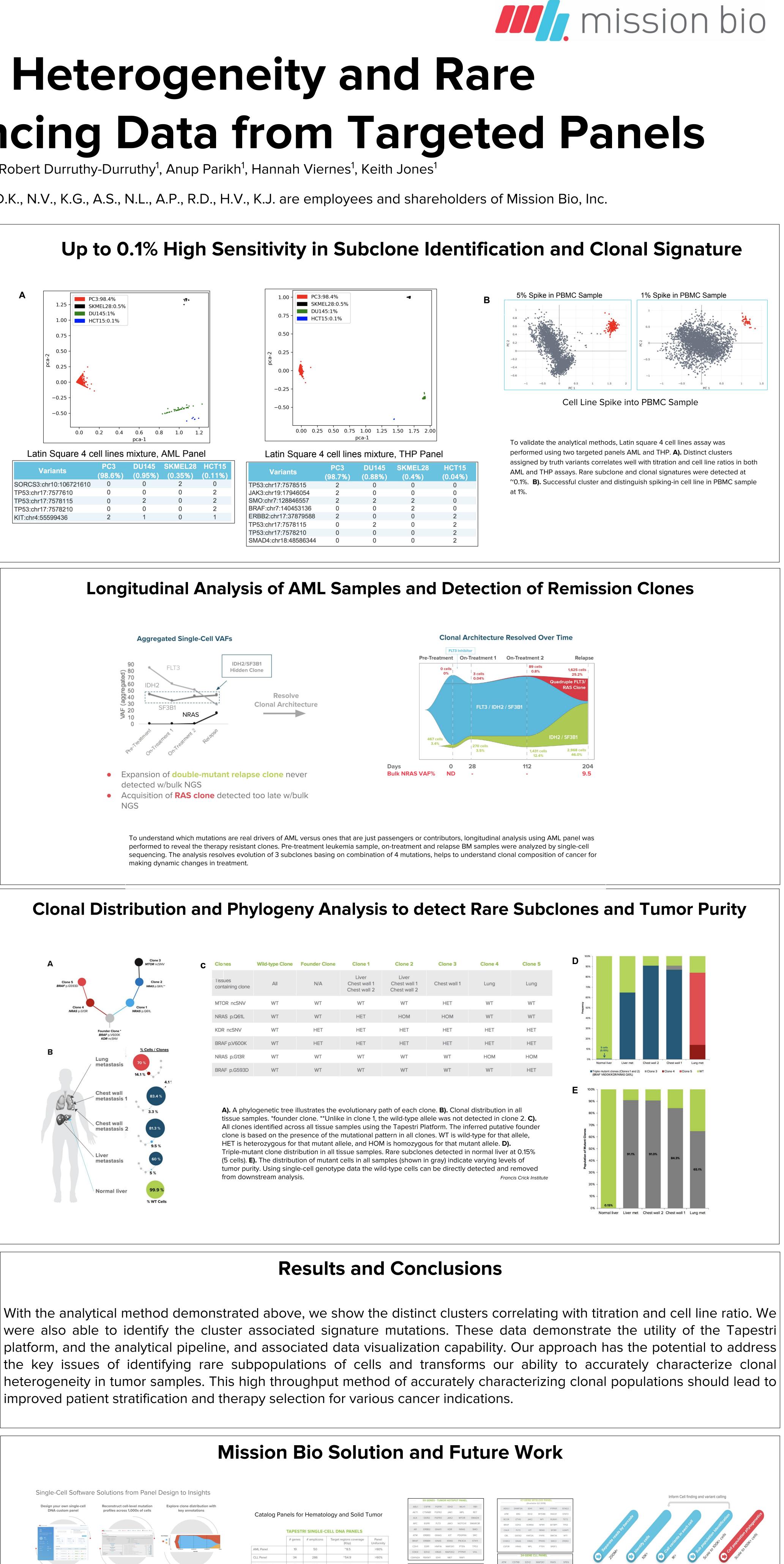
### Abstract

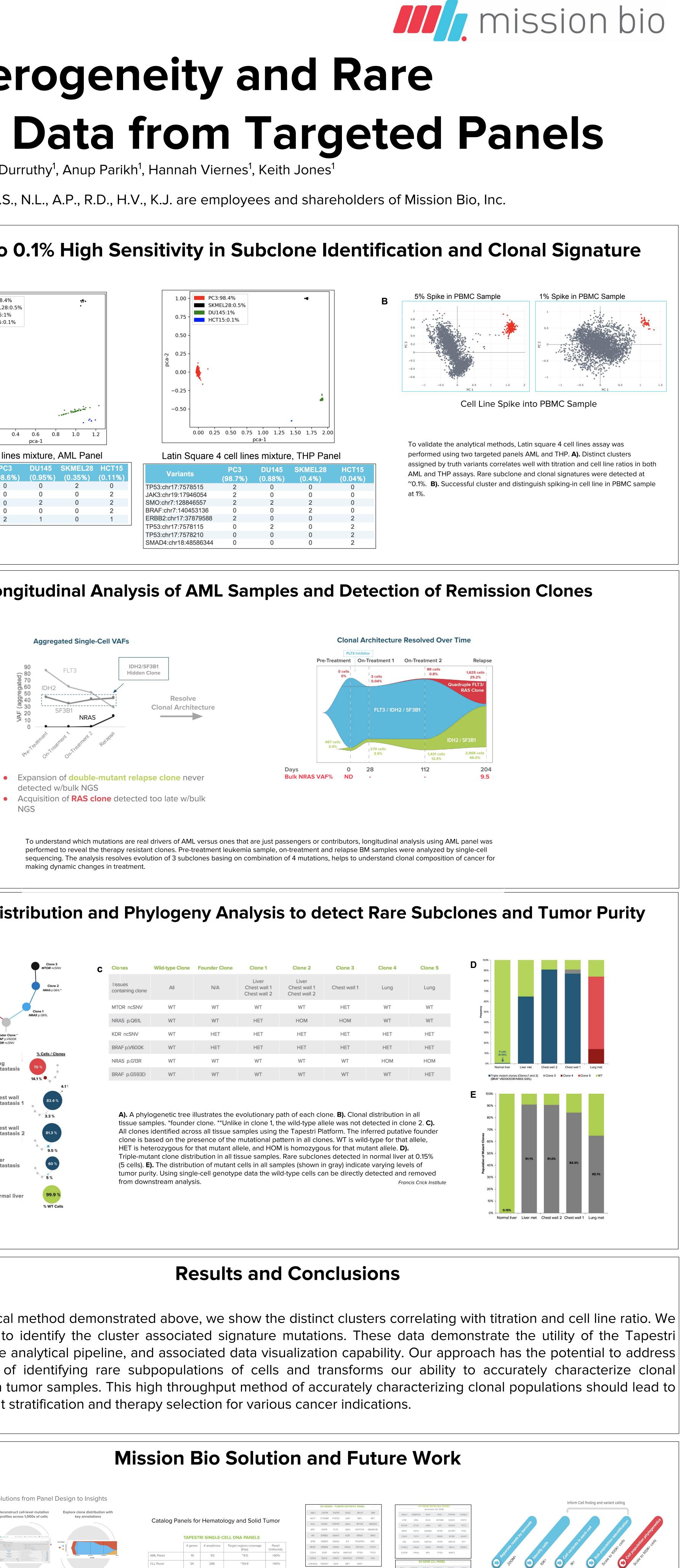
**Background:** With the advancements of single cell sequencing technologies it is now possible to interrogate thousands of cells in a single experiment. Single-cell RNA-Seq has been available for several years but high-throughput single-cell DNA analysis is in its infancy. Therefore, it is essential to develop new capabilities for assessing genetic variation present in rare cells and to better understand the role that these cells play in the evolution of tumor progression. To address these challenges and enable the characterization of genetic diversity in cancer cell populations, we developed a novel approach to identify mutation signatures which define subclones present in a tumor population.

**Methods:** Here we present subclone identification method using data generated on the Tapestri single-cell DNA platform and analyzed by Tapestri analytical workflow. The pipeline steps involve obtaining raw reads from the sequencer, removing adapters, aligning and mapping the reads, calling individual cells, and identifying genetic variants within each cell. After filtering for high quality variants, we then filter for data completeness to ensure only high quality data is used in downstream processing. The variant-cell matrix is then subjected to identification of subclones. Top variants defined the signature of each subclone are also identified. To validate our methodology, we used two different targeted sequencing panels on model systems with known truth mutations. Our pipeline shows the distinct clusters correlating with titration and cell line ratios. Cluster associated signature mutations were also identified. The pipeline can be used for multi sample analysis with time-series data from diagnosis to relapse or from primary site to metastasis to understand clonal diversity. These data demonstrate the utility of the Tapestri platform, the analytical pipeline, and associated data visualization capability. Our approach addresses key issues of identifying rare subpopulations of cells down to 0.1%, and transforms the ability to accurately characterize clonal heterogeneity in tumor samples. This high throughput method advances research efforts to improve patient stratification and therapy selection for various cancer indications.



Conflicts of interest: S.S., M.M., S.W., D.K., N.V., K.G., A.S., N.L., A.P., R.D., H.V., K.J. are employees and shareholders of Mission Bio, Inc.





		Cione 3 MTOR ncSNV	C Clones	Wild-type Clone	Founder Clone	Clone 1	Clone 2	Clone 3	Clone 4
Cione 5 BRAF p.G593D		Cione 2 NRAS p.Q61L**	Tissues containing clone	All	N/A	Liver Chest wall 1 Chest wall 2	Liver Chest wall 1 Chest wall 2	Chest wall 1	Lung
Clone 4 NRAS p.G13R		Cione 1	MTOR ncSNV	WT	WT	WT	WT	HET	WT
		NRAS p.Q61L	NRAS p.Q61L	WT	WT	HET	HOM	НОМ	WT
	Founder Clone * BRAF p.V600K		KDR ncSNV	WT	HET	HET	HET	HET	HET
	KDR ncSNV		BRAF p.V600K	WT	HET	HET	HET	HET	HET
	Lung	% Cells / Clones	NRAS p.G13R	WT	WT	WT	WT	WT	НОМ
	metastasis	70 %	BRAF p.G593D	WT	WT	WT	WT	WT	WT
	Chest wall metastasis 1 Chest wall metastasis 2 Liver metastasis	4.1 5 83.4 % 3.3 % 81.3 % 9.5 % 60 % 5 %	tissue sam All clones clone is ba HET is het Triple-mut (5 cells). <b>E</b> tumor puri	nples. *founde identified acro ased on the pr erozygous for ant clone distribu	r clone. **Unlik oss all tissue sa esence of the that mutant al ribution in all ti tion of mutant le-cell genotyp	te in clone 1, the second seco	bath of each clo he wild-type al the Tapestri Pl attern in all clor A is homozygo . Rare subclon nples (shown in Id-type cells ca	lele was not d atform. The in nes. WT is wild us for that mu es detected in n gray) indicat	etected in clor ferred putative I-type for that a tant allele. <b>D).</b> n normal liver a e varying level
	Normal liver	99.9 %							

improved patient stratification and therapy selection for various cancer indications.

el Design to Insights								
							59 G	
Explore clone distribution with						ABL1	CSF	
key annotations	Catalog Par	Catalog Panels for Hematology and Solid Tumor						
	Catalog r al		Tiemato	logy and Solia	lamoi	ALK	DDF	
						APC	EGF	
-		TAPESTRI SINGLE-CELL DNA PANELS						
Canad Phylograp		ATM	ERBI					
		# genes	# amplicons	Target regions coverage (Kbp)	Panel Uniformity	BRAF	ERBI	
	ANII Demol	19	50	~8.5		CDH1	ESF	
30 protestanent antestanent antestanent2 relajou Time Palint	AML Panel	19	50	8.5	>90%	CDK4	EZH	
	CLL Panel	34	286	~54.9	>90%	CDKN2/	A FBX\	
	Myeloid Panel	47	330	~66.0	>80%			
Tapestri	Tumor Hotspot Panel	59	244	~40.0	>80%	ASXL	.1 (	
Insights						DNMT:	ЗA	
						EZH2	2	
						FLT3	3	

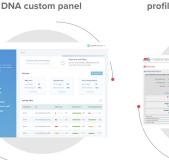
59 GENES - TUMOR HOTSPOT PANEL							
ABL1	CSF1R	FGFR1	IDH2	MLH1	RB1		
AKT1	CTNNB1	FGFR2	JAK1	MPL	RET		
ALK	DDR2	FGFR3	JAK2	MTOR	SMAD4		
APC	EGFR	FLT3	JAK3	NOTCH1	SMARCB1		
AR	ERBB2	GNA11	KDR	NRAS	SMO		
ATM	ERBB3 GNAQ KIT		PDGFRA	SRC			
BRAF	F ERBB4 GN		KRAS	<b>PIK3CA</b>	STK11		
CDH1	ESR1	HNF1A	MAP2K1	PTEN	TP53		
CDK4	EZH2 HRAS MA		MAP2K2	PTPN11	VHL		
CDKN2A	FBXW7	IDH1	MET	RAF1			
19-GENE AML PANEL							
ASXL1	GATA	42	KIT	PTPN11	TP53		
DNMT3A	IDH	1 K	RAS	RUNX1	U2AF1		
EZH2	IDH.	2 N	IPM1	SF3B1	WT1		
FLT3	JAK	2 N	IRAS	SRSF2			

CHD2 FAT1 MED12 PLCG2 CREBBP FBXW7 MYD88 POT1 XP

CXCR4 KLHL6 NFKBIE RPS15 ZM

DDX3X KRAS NOTCH1 SETD2

EGR2 LRP1B NRAS SF3B1



Multi-sample Analysis & Visualization

Primary Analysis