High-throughput Single-cell Targeted DNA Sequencing Using an Updated TapestriTM Platform Reveals **Rare Clones and Clonal Evolution for Multiple Blood Cancers**

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Abstract

The challenge in precision medicine has been improving the understanding of cancer heterogeneity and clonal evolution, which has major implications in targeted therapy selection and disease monitoring. However, current bulk sequencing methods are unable to unambiguously identify rare pathogenic or drug-resistant cell populations and determine whether mutations co-occur within the same cell. Single-cell sequencing has the potential to provide unique insights on the cellular and genetic composition, drivers, and signatures of cancer at unparalleled sensitivity. **Methods:** Previously we have developed a high-throughput single-cell DNA analysis platform (TapestriTM) that leverages droplet microfluidics and a multiplex-PCR based targeted DNA sequencing approach, and demonstrated the generation of high-resolution maps of clonal architecture from acute myeloid leukemia (AML) tumors. Here we present an update to the Tapestri Platform which employs new biochemistry and features improved firmware, software, workflow, and data analysis solution resulting in higher throughput, better sensitivity, specificity and unprecedented flexibility. Results: From cell prep to sequencing-ready libraries, the workflow can be completed within 2 days. We have validated the performance of an AML (19 genes, 50 amplicons) and a CLL (chronic lymphocytic leukemia) (34 genes, 286 amplicons) panel. We also developed a robust web-based design portal for custom targets. The updated biochemistry enables easy addition of new gene and loci targets into existing panels for improved coverage and updated studies. Using longitudinal AML and CLL samples, we were able to detect rare subclones of <0.1% prevalence, identify mutation co-occurrence, and characterize clonal evolution due to disease progression and drug treatment. **Conclusion:** We demonstrate that single-cell DNA sequencing can reveal the heterogeneity of blood cancers and map the clonal architecture and clonal evolution with higher sensitivity than bulk NGS methods. This is critical in patient stratification and drug selection over the entire course of treatment. Besides the catalog AML and CLL panels, the flexibility of system allows for analyzing SNV and indel mutations of any custom cancer DNA targets. Additionally, the system provides capabilities for quality control of gene edited cells, further advancing research into cancer therapies.





Single Cell DNA-Seq Data Analysis and Visualization RECENT ANALYSIS tapestri MDS1_tapestri-Insights-analysis MBM217.cells MBM231.cells VR15.cells RAJI MBM217.cells K362 MBM231.cells Fish Plots OPEN ANALYSIS N Variant & Cell Filtering Subclone Identification Variant Review Data Loading A significant challenge for high-throughput single cell sequencing is analyzing the complex data and visualizing results in biologically-meaningful way. We have updated the Tapestri pipeline and Tapestri Insight software, to more accurately and quickly reconstruct cell-level mutation profiles across thousands of cells, and explore clone distribution and progression with key annotations.

ulk Sequencing	Single-Cell Sequencing
ferred	Quantifiable
eterozygous ssumed)	Quantifiable
ferred	Quantifiable
ferred	Quantifiable
ndetectable	Detectable & Removable

barcoding to sequencing-ready libraries with

Panel ID	Species	Targets	# gene	# amplic on	Target region coverage (Kbp)	Mapped to Genome %
Testing Panel 1	Human	Gene exons, hotspot, ADOs	20	138	~24.2	97.7
Testing Panel 2	Human	Gene exons and hotspot	266	287	~53.0	91.9

To test the performances of the updated chemistry and designer pipeline, we designed three panels with 31, 138, and 287 (small, medium, large) amplicons respectively. The primer pools were pooled in equal volumes without any optimization, in order to simulate a standard custom panel at first pass (Mission Bio can also provide white-glove services to rescue specific designs and optimize individual amplicon performances after wet QC). Multiple runs were conducted for each panel with different cell types. An updated Tapestri analysis pipeline was used in analyzing the FASTQ data, collapsing cell barcodes, finding cells and calling genotypes. Typical cell outputs range from 5000 to 7,000 cells in these runs (30-40x sequencing depth), with <8% cell mixing (50/50 cell mix experiments) and <10% allele dropout rate. The run to run consistency data were shown above from multiple runs of Testing Panel 1. Bottom graphs show the panel uniformity for the runs and how the panel uniformity metrics were calculated and visualized.

High Sensitivity Detection of Rare Cell Populations

-0.50 0.0 0.2 0.4 0.6 0. pca-1		0.00 0.25 0	0.50 0.75 1.00 1.25 pca-1	•	Panel B Truth Variants TP53:chr17:7578515	PC3 2 2	DU145 0	SKMEL28	HCT15 0
Cell Percentages	PC3	DU145	SKMEL2 8	HCT15	SMO:chr7:128846557 BRAF:chr7:140453136	2	2	2	0
Designed Input Cell Composition	98.5%	1%	0.5%	0.1%	ERBB2:chr17:37879588	2	0	0	2
Panel A run results	98.6%	0.95%	0.35%	0.11%	TP53:chr17:7578210	0	0	0	2
Panel B run results	98.7%	0.88%	0.40%	0.04%	SMAD4:chr18:48586344	0	0	0	2

To test the sensitivity and specificity of the system, we mixed four types of cells at different concentrations (98.4%, 1%, 0.5%, and 0.1%), and run the mixed cell population on Tapestri with two targeted panels (Panel A-50 amplicons and Panel B-244 amplicons). The truth variant data from the four cell types were generated from bulk sequencing of pure cell lines, then used in Tapestri data analysis to determine cell types for all single cells from the Tapestri results. The resulted percentages matched closely to the desired cell input composition. Clones of 0.1% cells can be detected using both panels.

Panel A Truth Variants	PC3	DU145	SKMEL28	HCT15
SORCS3:chr10:106721610	0	0	2	0
TP53:chr17:7577610	0	0	0	2
TP53:chr17:7578115	0	2	0	2
TP53:chr17:7578210	0	0	0	2
KIT:chr4:55599436	2	1	0	1

FLT3

IDH2

 $\simeq 80$

멾 60

The Tapestri Precision Genomics Platform utilizes novel droplet microfluidics to access the genomic DNA. Together with updated chemistry and designer, it generates high coverage uniformity and low allele dropout for thousands of single cells, and enables detection of rare subclones down to 0.1%. Researchers now have a highly sensitive, targeted, and customizable solution for investigating genomic variation and clonal evolution in complex biological samples, as demonstrated in multiple AML and CLL longitudinal studies.

References:

Pellegrino M., et al., *Genome Research* (2018) McMahon C.M., et al., *submitted* (2019)

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Build Desired Targeted Panels ntuitive cloud-based Tapestri Designer this year to enable users to focus time and budget custom panels for their relevant targets, multiplexed at 20 to 500 amplicons. The targets can be available for AML, Myeloid, CLL, and Tumor Hotspot, to initiate the studies immediately or use as a base panel. For unique target sequences such as CRISPR edited cells, targets outside the exome, or for non-human animal models, expert bioinformatic support can be provided in custom project format. **Clonal Evolution in Longitudinal AML Samples** gh-throughput single-cell DNA sequencing of acute yeloid leukemia tumors with droplet microfluidics MDAnderson ancer Center Pellegrino, ^{1,5} Adam Sciambi, ^{1,5} Sebastian Treusch, ^{1,5} Durruthy-Durruthy,¹ Kaustubh Gokhale,¹ Jose Jacob,¹ Tina X. Chen, NRAS (G13R), ASXL1 (G646fs Geis,¹ William Oldham,¹ Jairo Matthews,² Hagop Kantarjian, ew Futreal.³ Kevur Patel.⁴ Keith W. Jones.¹ Koichi Takahashi.² Bio, Incorporated, South San Francisco, California 94080, USA; ²Department of Leukemia, ³Department of Geno, ⁴Department of Hematoporthology, The University of Texas MD Anderson Concer Center, Houston, Texas 77030 Single-cel 65 year-old male with AML Induction chemotherapy with cladribine and 20% -15% • Achieved complete remission (CR) at day 28 Relapsed 3 years after CR Pellegrino M., et al., Genome Research (2018) UCSF **Tapestri Reveals Therapy Resistant Clones** Penn **Clonal Architecture Resolved Over Time** Early identification of Gilteritinib sistant clones 28 days 84 days 92 days IDH2/SF3B1 Hidden Clone IDH2/SF3B1 Resolve **Clonal Architectur** IDH2/SF3B FLT3 **IDH2/SF3B** FLT3/NRAS 0 cells 3 cells 134 cells 1,625 cells McMahon C.M., et al., submitted (2019 Single-Cell DNA Sequencing Resolves the Genetic **Complexity Underlying CLL Progression** Clone 2 ATM W164X Het TM W164X Het тр1 → тр2 ATM p.V2757G 97.4% 2.6% 0% 95.8% 4.2% 0% ATM p.12888T 95.4% 4.6% 0% 78.6% 21.4% 0 In collaboration with Dr Esteban Braggio at Mayo Clinic, we have tested patient CLL samples at different time points. Results show that single cell VAFs are strongly correlated with bulk sequencing VAFs, but only single cell sequencing can resolve mutation zygosity and co-occurrence. Complex phylogenetic trees can be constructed from the single cell variant information to represent the clonal evolution (circle sizes correlate to subclone cell percentages)

Conclusions